# COMPASS<sup>®</sup> ENTEROCOCCUS AGAR

**ENUMERATION OF ENTEROCOCCI** 

### 1 INTENDED USE

COMPASS<sup>®</sup> Enterococcus Agar is a selective media used for the enumeration of enterococci in food and water.

#### 2 HISTORY

Enterococci are used as indicator organisms for fecal contamination and it has been proven that the majority of these bacteria produce a  $\beta$ -D glucosidase. The majority of media frequently used until today have contained esculin (6-7 dihydroxycoumarine-6-glucoside) which is hydrolyzed by the  $\beta$ -D glucosidase into two compounds : esculetin and glucose. Esculetin produces a brown component in presence of ferric ions present in the media, leading to a positive reaction with enterococci, but also with other unrelated secondary microorganisms. Indeed, a certain number of contaminating microorganisms can lead to a relatively high level of false positives, requiring a number of confirmation tests for all the colonies presenting a characteristic aspect.

The use of chromogenic and fluorgenic substrates for the detection of glucosidic activity, to specifically dectect enterococci, has been studied by numerous authors as Dufour in 1980, Littel & Hartman in 1983, Hernandez *et al.* in 1993. The association between X- $\beta$ -glucoside and the inhibiting nature of the formulation of COMPASS<sup>®</sup> *Enterococcus* Agar allows the direct enumeration by colony count of characteristic colonies, after only 24 hours of incubation and without confirmation.

#### 3 PRINCIPLES

Peptones, yeast extract and Tween 80 stimulate the growth of enterococci.

Yeast extract is a source of complex vitamin B.

Sodium chloride maintains the osmotic pressure.

The choice of an incubation temperature at 44 °C, associated with an judicious mixture of inhibitors, allows the high selectivity of the media and the inhibition of contaminating microflora.

The X- $\beta$ -glucoside insures the chromogenic revelation of the  $\beta$ -glucosidase activity of the enterococci. These colonies present a blue coloration after hydrolysis of the substrate (5-bromo-4-chloro-3-indolyl- $\beta$ -D-glucoside).

#### 4 TYPICAL COMPOSITION

The composition can be adjusted in order to achieve optimal performance.

For 1 liter of media :

- Peptones	27,5 g
- Yeast extract	5,0 g
- Sodium chloride	5,0 g
- Tween 80	1,0 g
- Inhibiting mixture	0,3 g
- X-β-glucoside	0,1 g
- Bacteriological agar	14,0 g

pH of the ready-to-use media at 25 °C :  $7,5 \pm 0,2$ .



#### 5 PREPARATION

- Dissolve 52,9 g of dehydrated media (BK183) in 1 liter of distilled or demineralized water.
- Stir slowly until complete dissolution.
- Divide into tubes or vials.
- Sterilize in an autoclave at 121°C for 15 minutes.
- Cool and maintain at 44-47 °C.
- For use in surface inoculation or membrane filtration, pour plates and let solidify on a cool, flat surface.

#### Use of the ready-to-melt media :

- Melt the media (if it is prepared in advance) or the ready-to-melt media (BM116) for the minimum amount for time required to achieve total liquefaction.
- Cool and maintain at 44-47 °C.

#### INSTRUCTIONS FOR USE 6

#### Food microbiology

- Transfer 1 mL of the sample suspension and its serial dilutions to empty, sterile Petri plates.
- Pour roughly 15 mL of molten media.
- Mix well by swirling and let solidify on a flat, cold surface.
- Incubate at  $(44 \pm 1)$  °C for  $(24 \pm 2)$  hours.

#### Water analysis

- Aseptically filter through a membrane a specific volume of sample water to test.
- To the surface of plates prepared as above, or to pre-poured media (BM157) brought to room temperature, place the membrane filtered side up and insure that the membrane and agar surface are in close contact.
- Incubate at  $(44 \pm 1)$  °C for  $(24 \pm 2)$  hours.

#### 7 RESULTS

The enterococci are characterized by blue to blue-green colonies on the agar.

See ANNEX 1 : PHOTO SUPPORT.

#### 8 **QUALITY CONTROL**

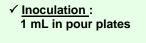
Dehydrated media : whitish powder, free-flowing and homogeneous. Prepared media : amber agar.

Typical culture response after 24 hours of incubation at 44 °C :

Microorganism	าร	Growth (Productivity Ratio : <i>P</i> <sub>R</sub> )	Characteristics
Enterococcus faecalis	WDCM 00087	$P_{R} \ge 50 \ \%$	Blue colonies
Enterococcus faecium	WDCM 00010	$P_{R} \ge 50 \ \%$	Blue colonies
Escherichia coli	WDCM 00013	Inhibited	-
Staphylococcus aureus	WDCM 00034	Inhibited	



✓ Reconstitution : 52,9 g/L ✓ Sterilization : 15 min at 121 °C



- ✓ Incubation : 24 ± 2 h at 44 ± 1 °C

**Dehydrated media**: 2-30 °C. **Ready-to-melt media in vials**: 2-8 °C. **Pre-poured media in plates**: 2-8 °C, shielded from light. The expiration dates are indicated on the labels.

Prepared media in tubes or vials (\*) : 180 days at 2-8 °C, shielded from light.
Prepared media in 55 mm Petri plates (\*) : 30 days at 2-8 °C, shielded from light.
(\*) Benchmark value determined under standard preparation conditions, following manufacturer's instructions.

### 10 PACKAGING

<b>Dehydrated media :</b> 500 g bottle	BK183HA
<b>Ready-to-melt media :</b> 10 x 100 mL	BM11608
Pre-poured media in Petri plates (Ø 55 mm) : 20 plates	BM15708

### 11 BIBLIOGRAPHY

Microbiolgy, Washington, D.C., USA. Abstr. Q-69 : 205.

Littel, K.J., and Hartmann P.A.. 1983. Fluorogenic selective and differential medium for isolation of fecal streptococci. Applied and Environmental Microbiology, **45(2)** : 622-627.

Hernandez, J.F., Pourcher, A.M., Delattre, J.M., Oger, C., and Loeuillard J.L.. 1993. MPN Miniaturized procedure for the enumeration of faecal enterococci in fresh and marine waters, The must procedure. Water Research, **27** : 597-606.

#### 12 ADDITIONAL INFORMATION

The information provided on the labels take precedence over the formulations or instructions described in this document and are susceptible to modification at any time, without warning.

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## **ANNEX 1 : PHOTO SUPPORT**

# COMPASS<sup>®</sup> Enterococcus Agar

Enumeration of *Enterococcus* spp.

### **Results :**

Growth obtained after 24 hours of incubation at 44 °C (pour plate inoculation).

Enterococcus faecalis

Colony characteristic : blue color (hydrolysis of X-β-D-glucoside)

